

Can gene editing provide solutions to plant pathogen sensitivity ?

Erik Andreasson
SLU Alnarp
Resistance biology

Acknowledgements

Awais Zahid

Nam Kieu Phuong

Marit Lenman

Svante Resjö

Naga Charan

Milla Karlsson

Frida Meijer Carlsen (KU)

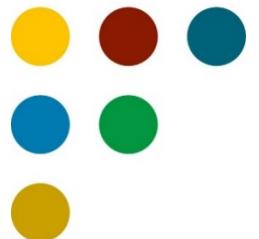
Bent L Petersen (KU)

All others at the resistance biology unit



novo
nordisk
fonden

FORMAS



SCIENCE AND
EDUCATION
FOR
SUSTAINABLE
LIFE

Resistance genes (R-genes) – make the plants more resistant

Reception of the pathogen

Co-evolution with pathogens can give resistance loss

For many pathogens no corresponding R genes are known

Susceptibility genes (S-genes) -make the plant more sensitive to infection

Negative regulation of defence (often salicylic acid (SA) related)

Transport of nutrients to pathogens

Inactivation gives recessive resistance

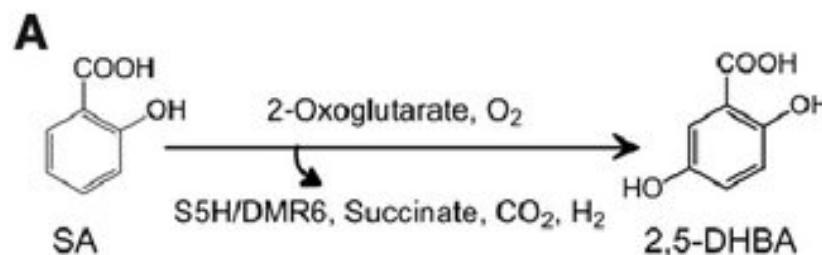
Genomeditering (CRISPR/Cas9) or looking for mutants

Negative effects by mutations?

Function of DMR6 –A known sensitivity gene

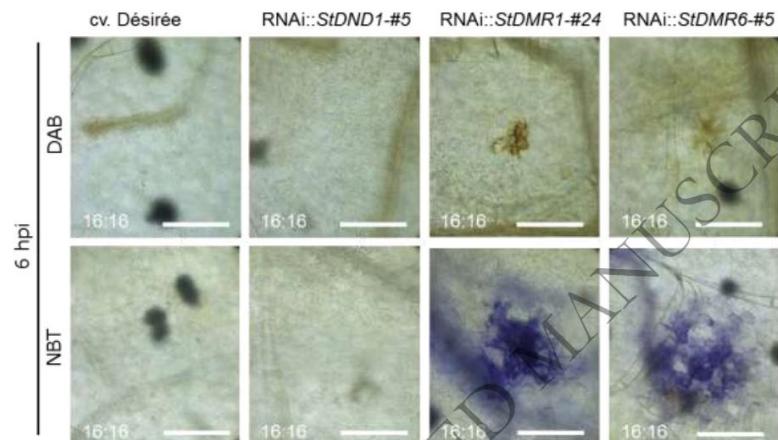
- DMR6 catalyze the conversion of SA to 2,5-DHBA on Arabidopsis and Tomato

(Zang et al. 2017; Thomazella DPT et. al. 2021)

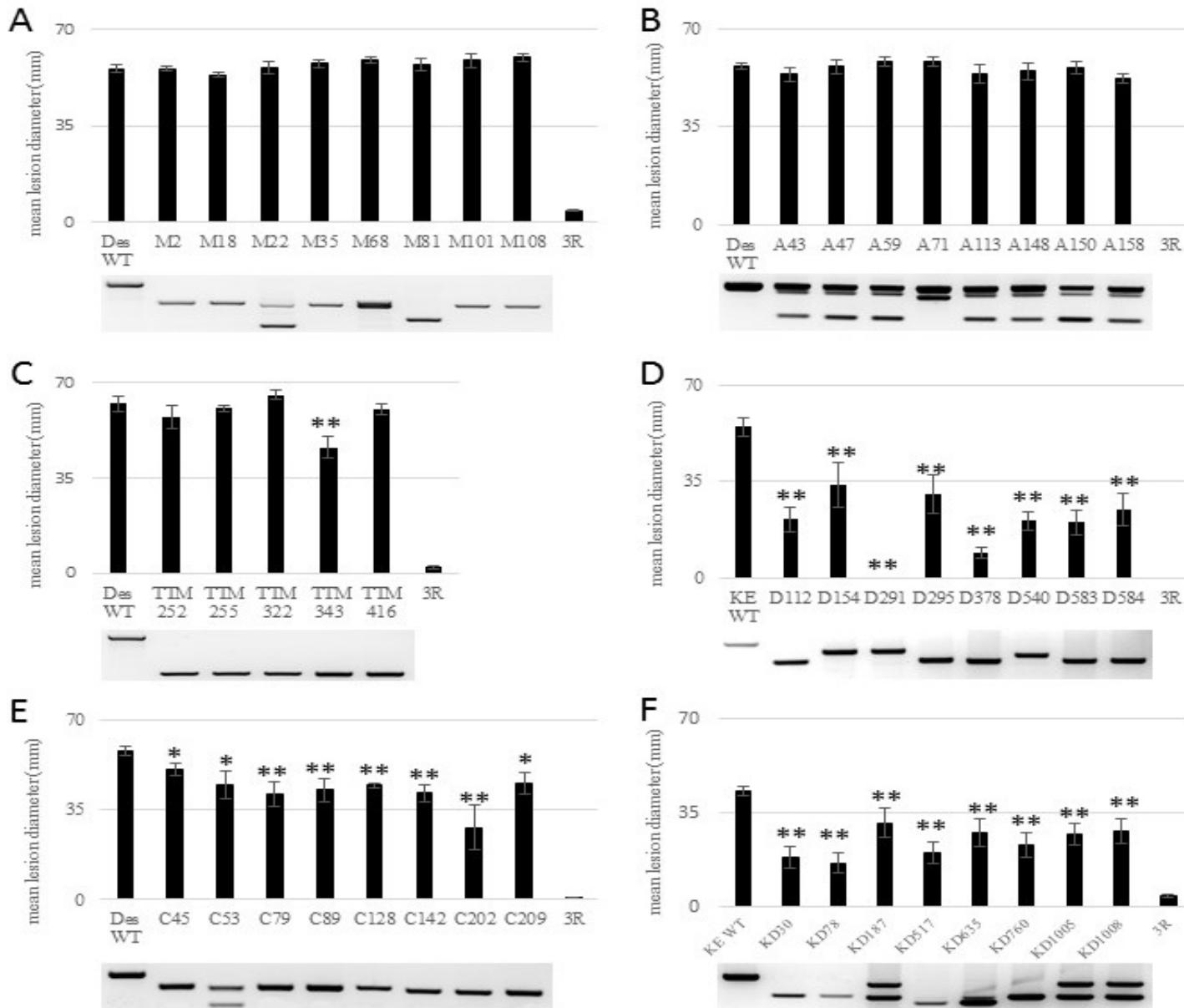


- StDMR6-silenced plants display early ROS accumulation after late blight attack

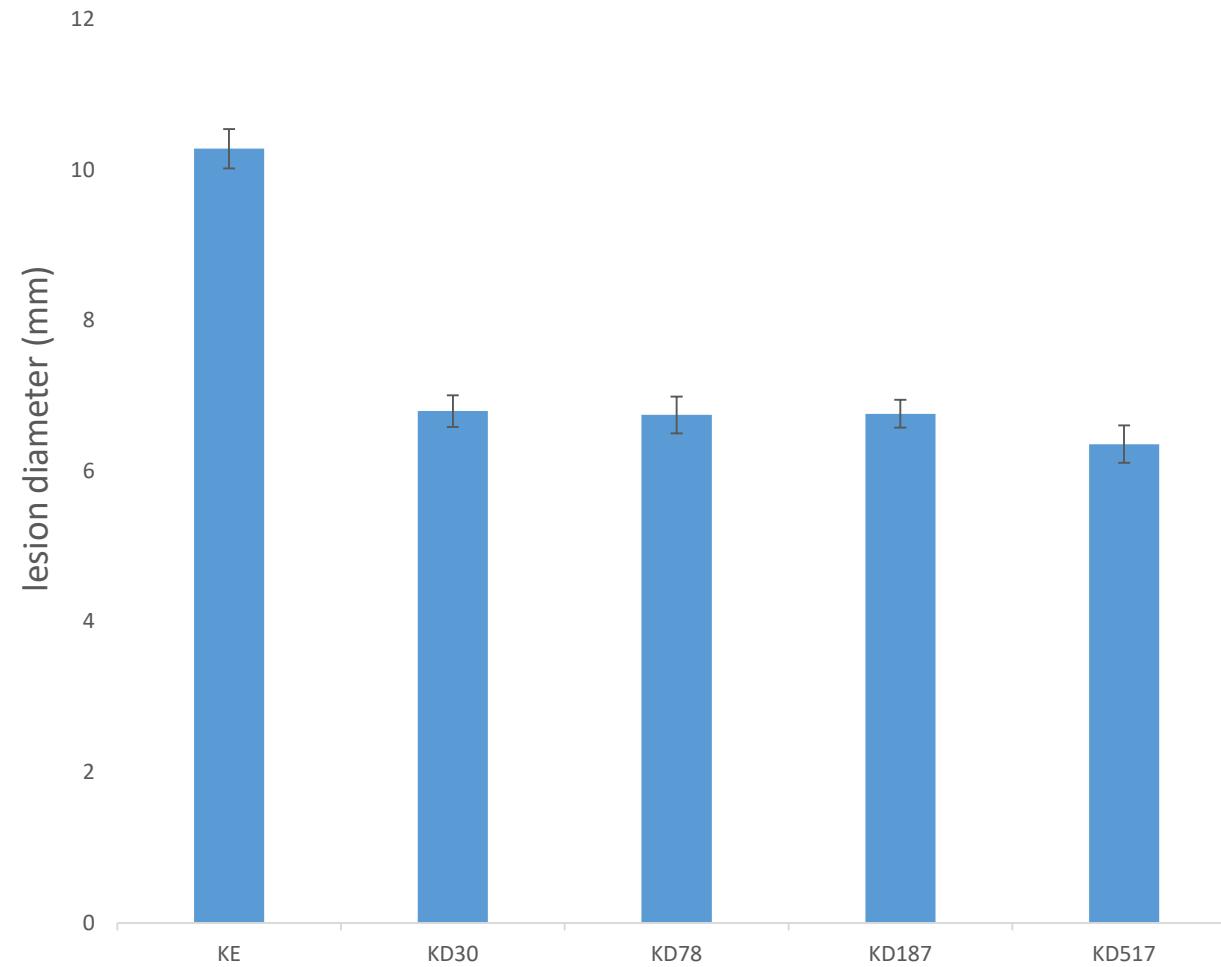
(Sun K et. al. Manuscript 2022)



Screening of CRISPR/Cas9 generated S gene deletions in King Edward potato by lesion diameter of late blight

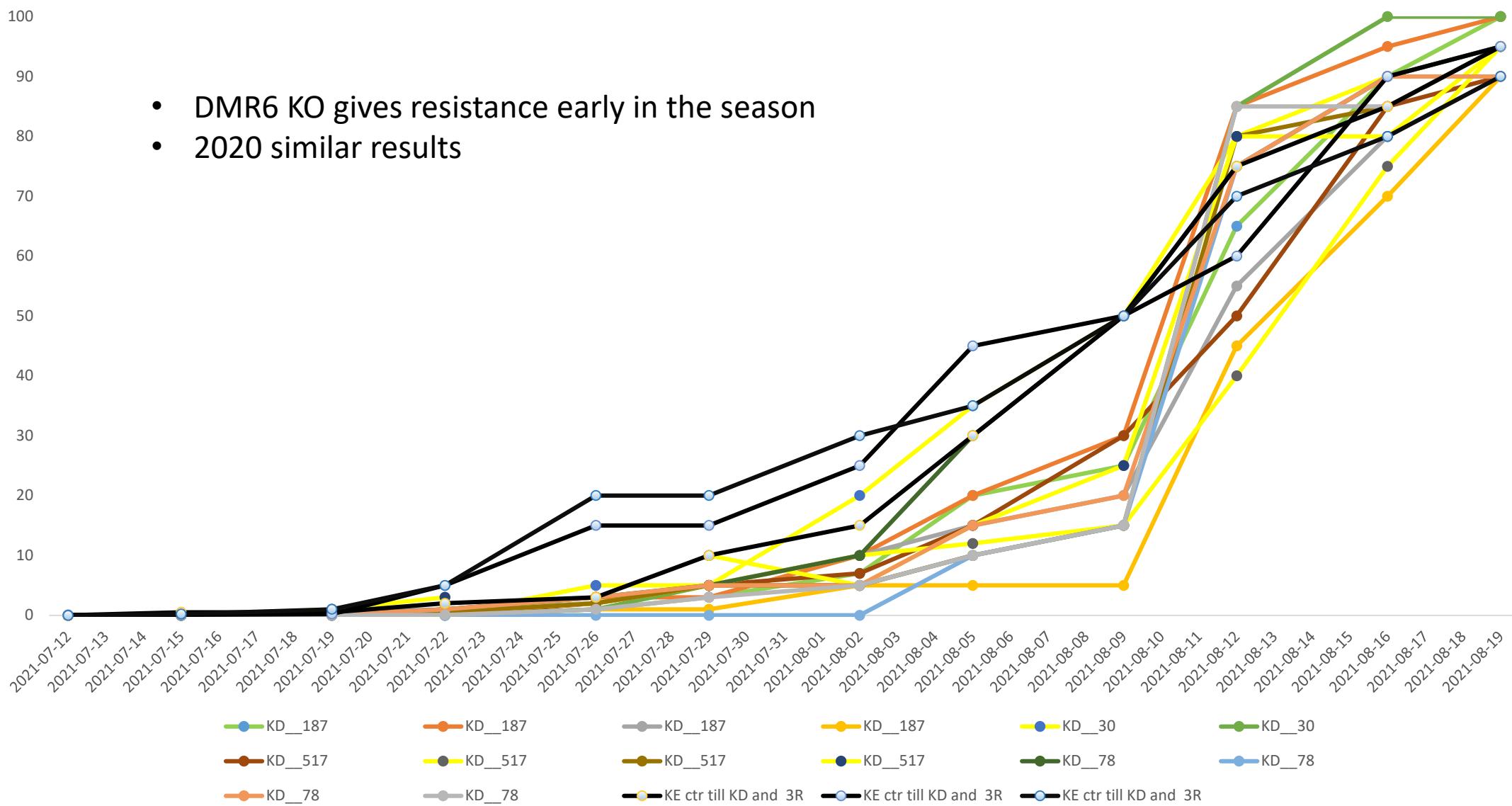


DMR6 KO (KD) result in less Early blight (*Alternaria solani*) in controlled conditions



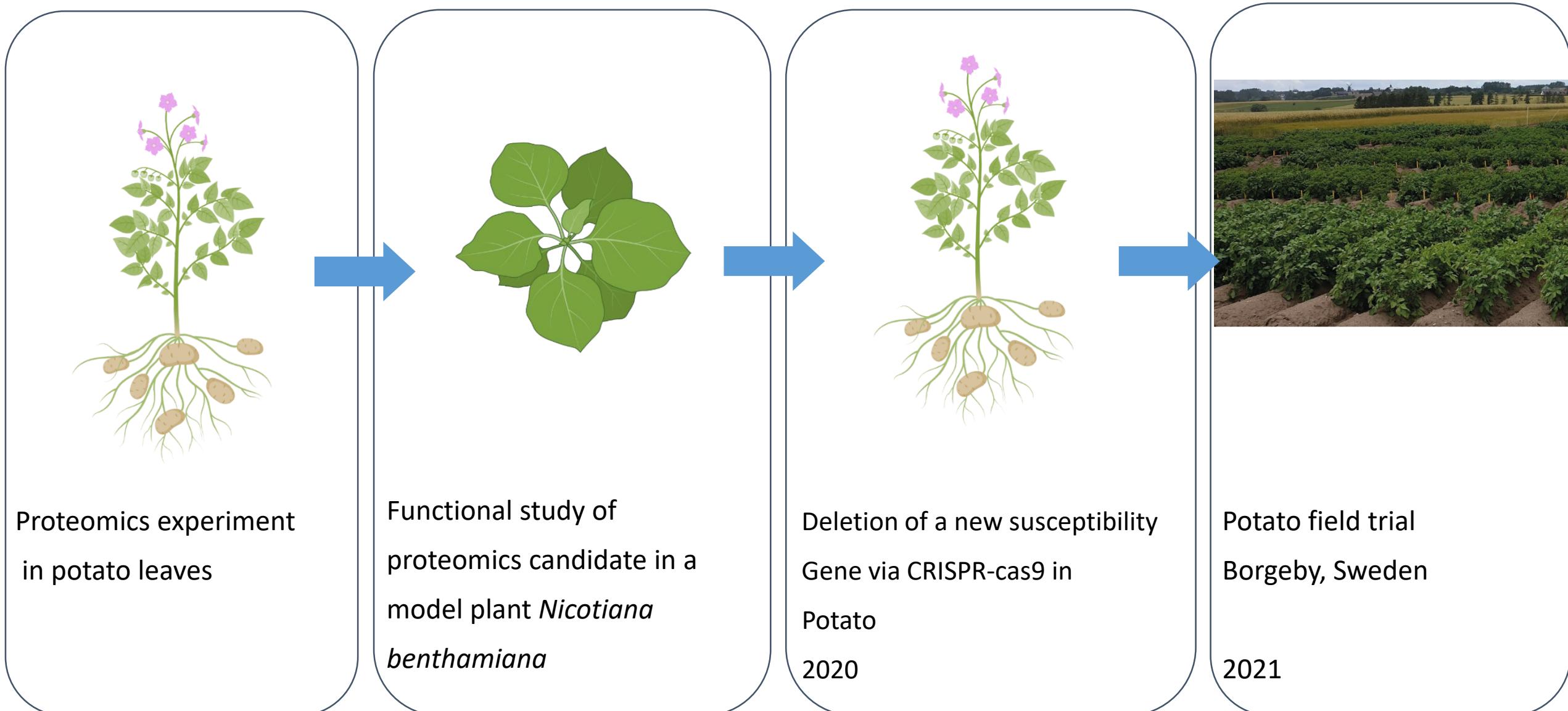
Late blight scoring of field trials in DMR6 CRISPR KO King Edward potato 2021 (black are controls)

- DMR6 KO gives resistance early in the season
 - 2020 similar results



New susceptibility gene -From laboratory to field

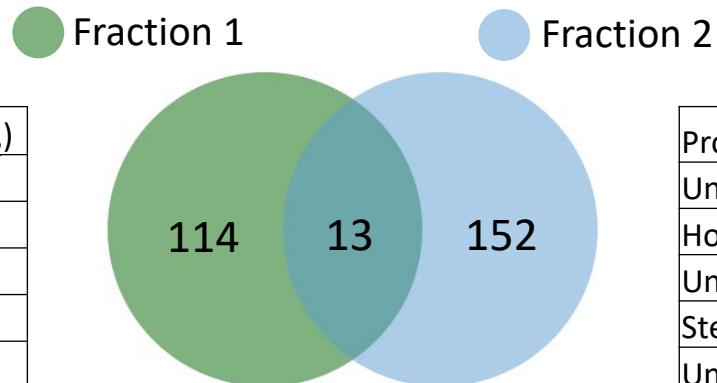
Awais Zahid



Potato proteomics after activation of immunity reactions

Fraction 1

Protein Names	Fold change (\log_2)
Mitochondrial SBP40	4.0
SRC2-like protein	3.6
Rop guanine nucleotide exchange factor	3.4
Sucrose synthase	3.2
Alliin lyase	3.2
CC-NBS-LRR protein	3.2
Uncharacterized protein	3.1
30S ribosomal protein S8	2.5
Integrase core domain	2.3
PSI-H	2.2
Chlorophyll a-b binding protein (LHCB3)	-5.6
ATP synthase subunit c	-2.8
Hydrolase	-2.2
Photosystem II reaction (PSII-L)	-2.0
GDP-mannose 4,6-dehydratase	-1.9
32 kDa thylakoid membrane protein	-1.8
Thioredoxin-dependent peroxiredoxin	-1.7
Chlorophyll a-b binding protein	-1.7
NAD(P)H-quinone oxidoreductase	-1.6
Uncharacterized protein	-1.6



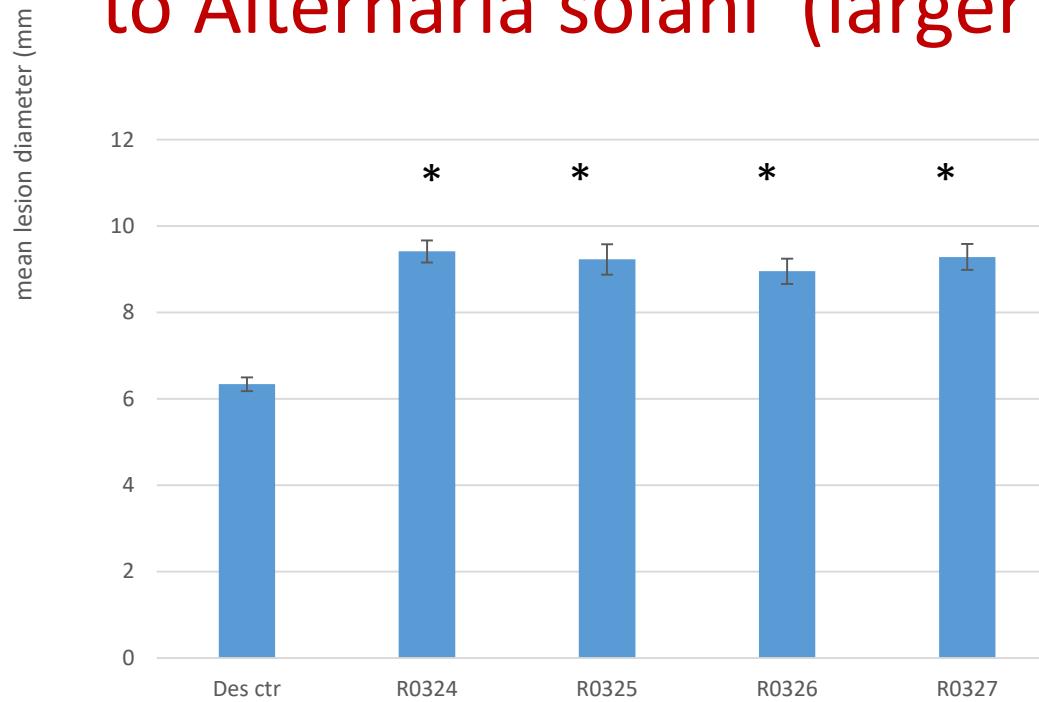
Fraction 2

Protein names	Fold change (\log_2)
Uncharacterized protein	5.6
Homeobox-leucine zipper protein	4.7
Uncharacterized protein	4.1
Stem 28 kDa glycoprotein	3.3
Uncharacterized protein	3.2
PS II reaction center protein L	3.0
3-isopropylmalate dehydratase	2.9
4F5 protein family protein	2.8
4F5 domain-containing protein	2.8
Plastocyanin	2.8
5'-3' exoribonuclease	-6.2
NAD_Gly3P_dh_N	-4.6
Mitogen-activated protein kinase	-2.5
Chlorophyll a-b binding protein	-2.4
Histone H2B	-2.3
BZIP domain-containing protein	-2.2
Hexose transporter	-2.1
3-oxoacyl-CoA synthase	-2.0
Poly(RC)-binding protein	-2.0
Pre-mRNA-splicing factor cwc23	-1.9

New S genes and functions

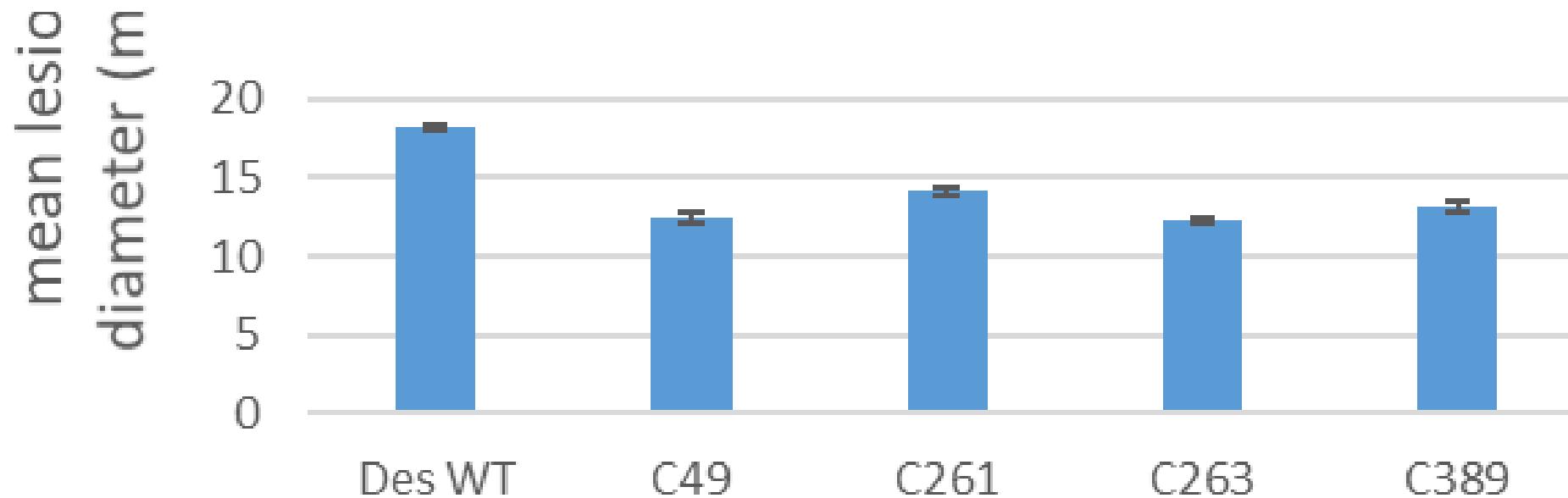
- DMR6 affects salicylic acid degradation, and have stress markers activated without stress
- One (SAs1) in the apoplast (unpublished, only Alternaria?)
- One (SPi1) in the thylakoid lumen (patent application) Broad spectrum resistance and in crops and also salt stress

Potato overexpressing SAs1 have a susceptible phenotype to *Alternaria solani* (larger lesions)



Des WT R0324
Leaf no.3 (bottom to top)

CRISPR-SAs1 Knock out Potato plants inoculated with *Alternaria solani* are more resistant to Early blight (torrfläckssjuka)

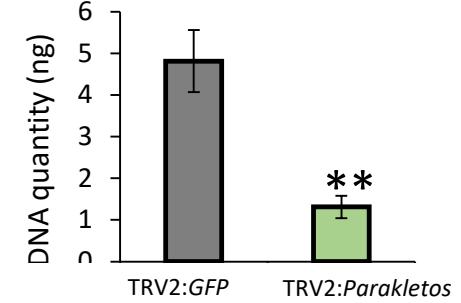
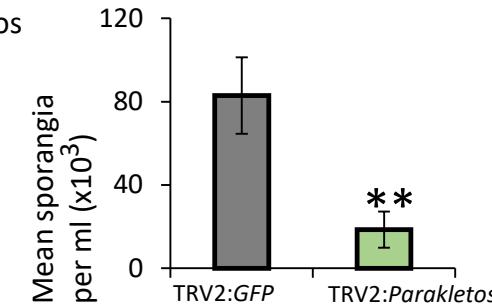
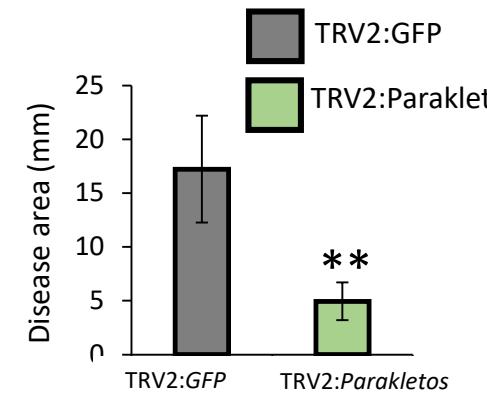


Functional test of candidate proteins reveals *Spi1*

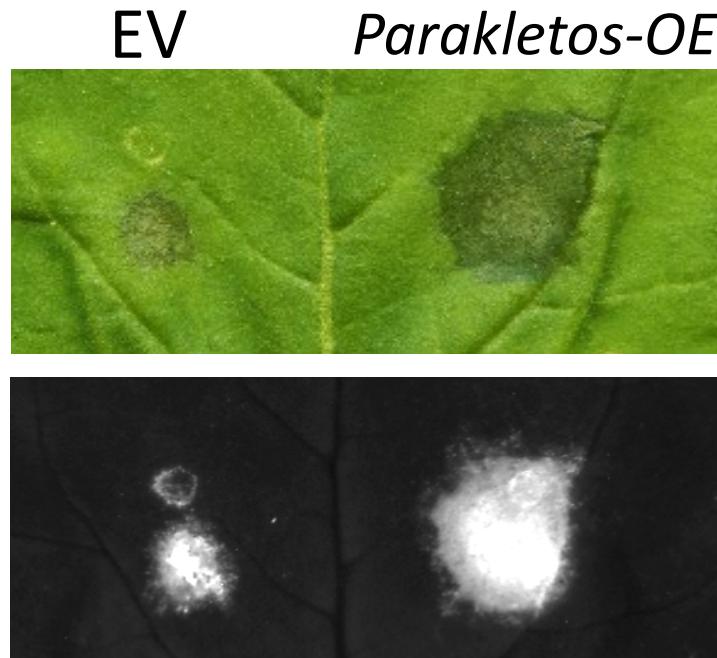
TRV2:*GFP*



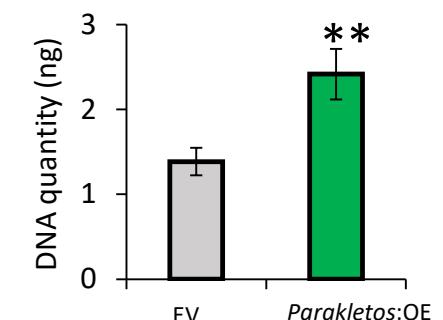
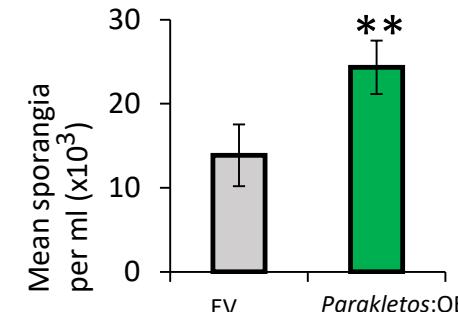
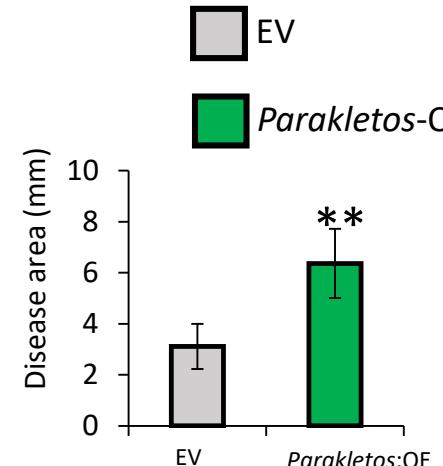
TRV2:*Parakletos*



EV

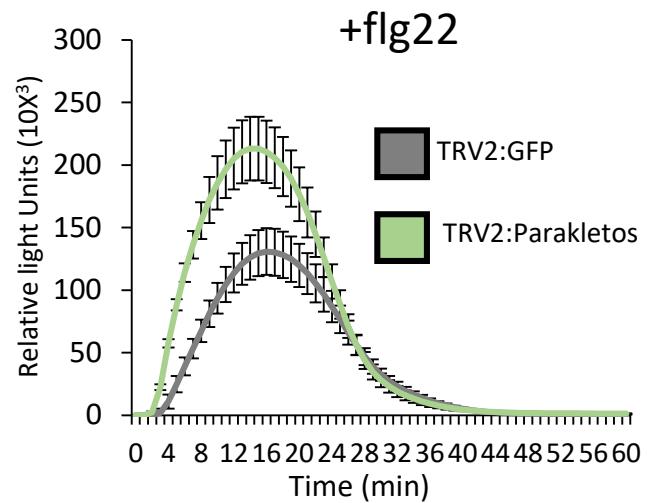


Parakletos-OE

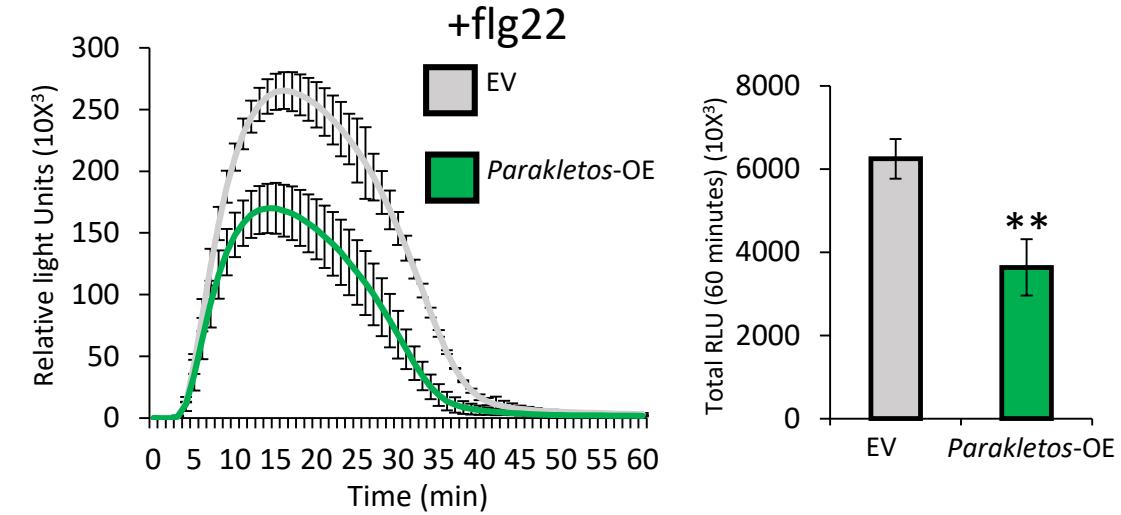
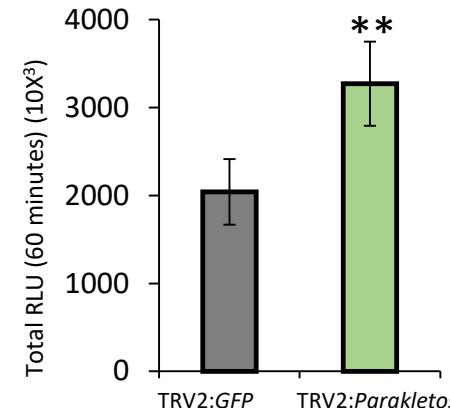


**Parakletos* negatively involves in plant immunity

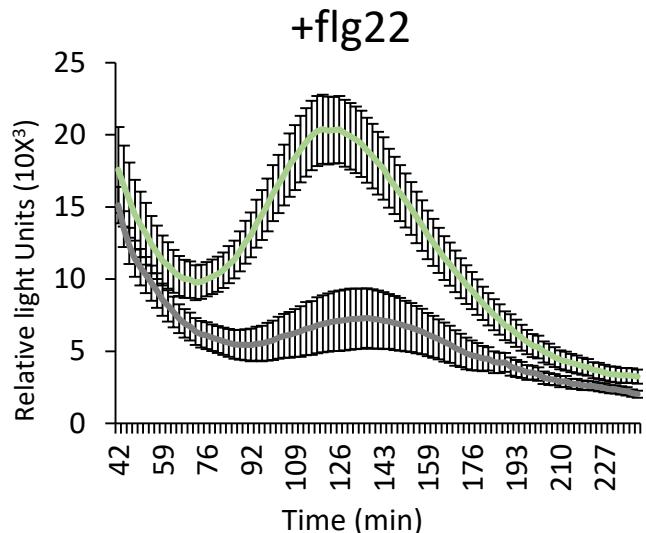
Spi1 suppresses ROS burst



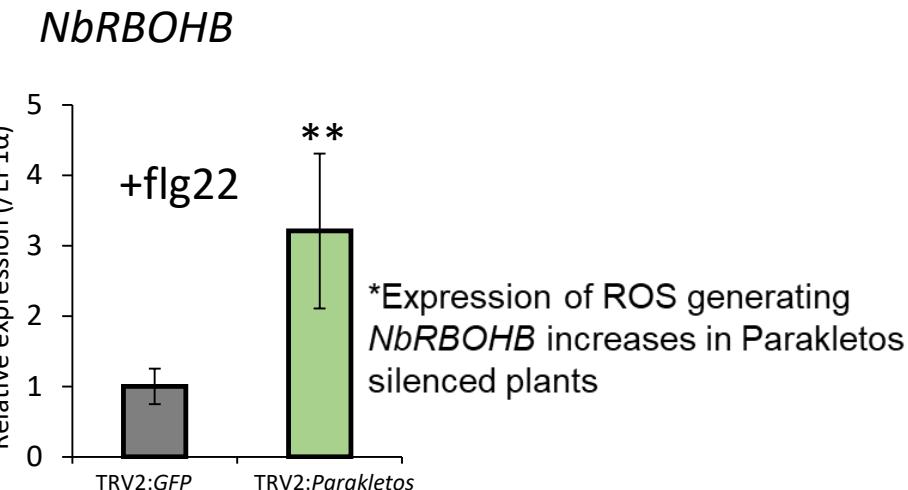
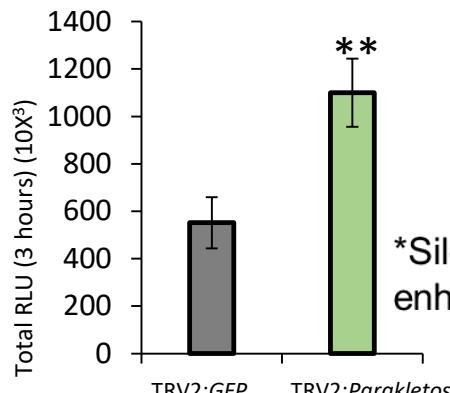
*Silencing of Parakletos enhances 1st ROS burst



*Parakletos-OE suppresses 1st ROS burst



*Silencing of Parakletos enhances 2nd ROS burst



*Expression of ROS generating NbRBOHB increases in Parakletos silenced plants

Summary

- Gene editing by CRISPR/Cas9 of potato is efficient
- Increased resistance to pathogens and salt have been achieved by inactivation of S genes (susceptibility genes)
- So far no negative agricultural effects of mutation of these S genes has been detected in potato